Volume 6: Issue 1: 2022

Review article

https://doi.org/10.56770/jcp2022615

SCN1A GENE MUTATION; A RISING CAUSE OF HUMAN EPILEPSY SYNDROME

Muhammad Akram Choohan¹*, Ishtiaq Ahmed², Shahzada Khurram Syed³, Muhammad Naeem⁴

¹University College of Conventional Medicine, The Islamia University of Bahawalpur, Pakistan ²School of Medical Science, Gold Coast Campus, Griffith University, Southport, QLD, Australia ³School of Health Sciences, University of Management and Technology, Lahore, Pakistan ⁴Department of Pharmacy, Shah Abdul Latif University Khairpur, Sindh.

Submitted 1st November 2021, Accepted 7th March 2022

ABSTRACT

Epilepsy is found to be very common neurological disorder; it covers a wide range of abnormalities lying within brain. Generation of electrical activity of brain that is action potential and then its proper propagation in central nervous system (CNS) and Peripheral Nervous System (PNS) upto the target site is mandatory for the proper functioning of brain. The human sodium channel (SCN) family plays its critical role in all these neuronal mechanisms. These channels are prone to number of channelopatheis and mutations that are the key point of research and discussion now a day. About 700 mutations of *SCNIA* gene have been identified. This gene is considered as the commonly mutated gene in human epilepsy. This review is on the structural, functional and pathological aspects of SCN1A gene and its associated channelopathies. This will explore the advances in epilepsy genetics and may help in designing for new therapeutic modalities for treating epileptic patients.

Keywords: SCN1A gene, Epilepsy, Mutation, Dravet syndrome

*Corresponding Author. E-mail: choohansb@gmail.com

INTRODUCTION

Epilepsy is a chronic syndrome associated with other brain diseases. It is actually a collection of diverse neurological disorders which involve the unprovoked seizures. These unprovoked seizures are the sign of abnormal electrical activity in the CNS. Epilepsy is long-lasting illness that upsets the whole life of patient and make it dependent on medications for long time [1]. Seizures associated with epilepsy may come in several ways. It majorly depends upon the area of the brain that is affected and also on the age of the person [2]. The most common type of seizures are convulsive seizures and they account for (60%) of total seizures. One third generalized seizures are due to the involvement of both hemispheres at the starting phase of epilepsy. Only one hemisphere is involved in two third of focal seizures which may convert into generalized seizures later. Other 40% of seizures are non-convulsive. Acquired epilepsy is about 20-30 %, causes may be head injury, brain tumor, toxicity of any chemical, drugs, genetic, anxiety, depression

and hypoglycemia or any other cause. The remaining epilepsy about 70-80 % is admitted as due to genetic factor [3]. The base of human epilepsy is human brain abnormality. It is due to the generation of abnormal impulses in the brain that majorly results from genetic mutations [4]. Nearly 40% cause of this disease is unidentified [5]. Unprovoked seizures are idiopathic. Idiopathic epilepsies are usually caused by genetic variations. Number of mutations may get responsible for these genetic variations in many unexplained genes [6]. Epilepsy becomes more common with increasing age **[7].** There are about 10% chances for all people to have a seizure near the age of 80 years [8]. In developed countries, infants and elderly people are more to get epilepsy while in developing countries the scenario is opposite. This is because of changes in the frequency of underlying causes [9]. About 4% of people worldwide (65 million) have epilepsy [10]. Each twenty eighth woman and twenty first man is getting disease of epilepsy [2].



Figure 1: Country wise specific epidemiology/prevalence

DRAVET SYNDROME

It is disastrous babyhood epilepsy illness in which the seizures are usually related to intelligence debility and not responding to medicines. Dravet Syndrome contains marginal and conventional DS, wherein only a lot of clinical features are seen in the patients **[11, 12, 13]**. DS is considered as general status epilepticus starting at the age of 6 month approximately, with other types of seizures, including myoclonic, partial, absence, and atonic seizures happening after passing the time of one year. In traditional DS, progress is postponed and patients every so often hurt from motor decline, as well as ataxia and spasticity **[14]**.

Dravet syndrome (DS) has categorized by persistent seizures in the 1st year of an infant. DS develops to further types seizure like myoclonic and focal seizures, psychomotor delay, and ataxia **[15]**. It is categorized by intellectual damage, behavior disorders, and motor deficits. It is also related with sleep disorders **[16]**. The seizures beard by persons with Dravet syndrome convert into worst as there is no prediction of disease with the age of patient when first detected. This joined with the kind of severity different between each person detected and the seizures become drugs resistant made a challenge to improve treatments **[16]**.

GENETICS

It is analysed that epilepsy is caused due to abnormalities in number of unknown genes. These abnormalities most probably include gene mutations. Research has shown that these mutations are most frequently single gene mutations [17]. In the majority of cases genes that are involved in epilepsy are those genes involved in ion channels. Although, the underlying defective mechanisms leading to epilepsy is not well comprehended [18]. The electrical excitability is due to these channels. It looks that epilepsy is due to variation in the voltage gated sodium channels. Action potential of membrane motivates the channel and a change is done that depends upon voltage. Penetrability of sodium ions is amplified by the change in depolarization of membrane that outcomes in the more depolarization in the entire cell [19]. After this depolarization ends in the inactivation and in the reaction of depolarization channel closed and penetrability to sodium ion is declined that outcomes in the rebuilding of the resting potential status. α subunit a highly treated nearly 260-kDa preparing sodium channel protein, that covers of four uniform and even domain named I-IV and each domain is needing six trans membrane parts recognized as S1-S6 [20].

Between the S5 and S6 a P-loop like hairpin sandwiched making a part of the channel pore and interrelating domain III and IV. An initiation door way is made by the formation of loop in the cell [21]. Channels are connected in the mature central nervous system the channels are linked by $\beta 1,\beta 3$ and B2 B4 subunits. That are appeared in parallel style. so as the α subunits are related with the β 1 or β 3 and $\beta 2$ or $\beta 4$ [11, 12, 13]. Each β subunit has a lone Trans membrane portion, an extracellular loop made of IgG-like and a C-terminus inside the cell. A subunits are bounded with $\beta 2$ or $\beta 4$ subunits via disulfide bond [14]. Non-covalently β 1 or β 3 subunits are linked with α subunit. The communication of β subunits with α subunit results in variation that make the α subunit kinetics, voltage and localization dependent. Voltage-gated Sodium channel α subunit are coded by 9 genes in animal and human beings such as SCN1A, while SCN11A (SCN6A and SCN7A) presenting the same gene that encodes for the sodium non voltage-gated channel. Nine dissimilar isoforms are encoded by all these genes recognized as Nav 1.1 done withNav1.9 [22, 23].

In CNS 4 isoforms shows them at higher stages (Nav1.1, Nav1.2, Nav1.3 and Nav1.6) and (Nav1.6, Nav1.7, Nav1.8 and Nav1.9) 4 isoforms show them in PNS at higher stages elevated. Firstly (Nav1.4) in grown persons skeletal muscle (Nav1.5) in developing heart muscles and the residual 2 isoforms are appeared. In the course of the growth of CNS these isoforms are founded in miscellaneous sites [24]. In the natal age Nav1.1 be an indication and grows up to childhood [25]. There may be a stimulus of chemical or mechanical is changed into electrical impulses in the cells in their excited state via voltage-gated sodium channels [26]. The formation of these channels by useful pore-farming α subunits and supplementary β subunits. Until now, 9 unlike forms of genes SCN (1-9A) are documented to encode the 9 unlike Na channels. Epilepsy related to inactivation of SCN2A and SCN1A [27] genes, SCN4A gene is related to myotonia disease, SCN5A gene is related to Brugada syndrome (OMIM 601144), young patients suffering with cardiac arrhythmias. Ataxia and cerebellar atrophy are disorders related to SCN8A gene. SCN9A gene is related to activating mutations and inactivating mutations unrelated with pain, with differing phenotype, hyperalgesia [28].

In the gestation period of fetal growth Nav1.2 acquires major level and in the maturity got at topmost level [25]. Nav 1.3 acquires its topmost level of appearance at natal phase and generally undetectable in entirely grown individuals. Therefor it is declared as fetal isoforms [25] but it can be founded at lowest level of human beings maturity [29]. Nav 1.3 is understood in NS of the matured rat afterward ANS damage of their dorsal root ganglion [30, 31] and in few rodents with epilepsy models [32, 33]. Nav 1.6 displays that one appearance in the late fetal growth, initial perinatal [34] and adulthood phases [35]. Nav 1.1 channel is founded abundantly in the area of spinal cord and matured CNS containing caudal reign while in rostral regions, positions of Nav 1.2 are full [36]. Nav 1.1 is founded in the cell bodies and dendrites up to highest level [37, 38]. It is mostly founded in the initial portion of axon for fast awkward parvalbumin positive neurons [39]. Nav 1.2 abundantly existed in the dendrites and axons without myelin sheath [37]. Nav 1.6 is existed in the sensory and motor

pathways and their sub-cellular dispersals existed in dendrites, axons, and cell bodies and post and presynaptic positions [40, 41, 42]. Nav 1.6 is prime sodium channel at node of Ranvier [42, 43].

With their unlike sub-cellular dispersal and functional characteristics, each isoform for sodium channel is usually doing a vital job for beginning and transmission of action potential in membrane. For example, differentiating in matching to the earliest corner action potentials are motivated in the early portions. Due to effect of this propensity travel towards far most end is Nav 1.6 channels with extraordinary concentration in contrast to the amplified concentration of Nav 1.2 in the initial end, little beginning is for Nav 1.6 channels start [44]. Early portion of parvalbumin-positive basket cells of axon wherever Nav 1.1 channels are existed in the clustered form [39]. The completion portion of axon axle of parvalbumin -positive basket cells are scheduled nearby the activated neurons physically and early dendrites. The main task is to regulate the reactivity of all connection.

SCN1A

Humans protein which encodes by SCN1A gene (Sodium channel voltage-gated) is symbolized as Nav1.1, type I, asubunit (SCN1A), Nav1.1 is named as neuronal voltage-gated sodium-channel also. To explain the concept of diseases with seizures and their reasons related to SCN1A the term is known as "channelopathies". Neuronal dysfunction is due to the molecular anomalies which in resulted in excitement of cortical network. SCN1A is an important part of gene group that encodes for Na channel. The location of this gene is 2q24 and making a cluster including SCN2A & SCN3A [45]. Na- channel with Alpha subunit participate in the pore formation of membrane. A subunit having four domains with six transmembrane segments related with loops. S5 & S6 contains the remaining of pore lining and the P-loop which is discussed above related to S5 & S6. Residuum with positive charge which can be detected by voltage detector [15]. In all the segments of Nav1.1 having pathogenic variants which are related to epilepsy, they present up to in N-terminus also, in the voltage measuring device, the p-loops in D1-D5 and excessively present in C-terminus [46, 47].

Here are two kinds of epilepsy syndromes recognized. One is Generalized Epilepsy with Febrile Seizures Plus (GEFS+).and the other is extreme epilepsy of childhood, known as Dravet Syndrome or Severe Myoclonic Epilepsy of Infancy (SMEI) are in the results of mutations in SCN1A gene. The attacks (seizures) with pyrexia are known as GEFS+ is an autosomal (milder) highest hereditary epilepsy syndrome. The GEFS+ with unlike kinds of its configuration in uninfluenced carriers, simple Febrile Seizures for most of patients, Febrile Seizures Plus and infrequently ample painful epilepsy. By the statements of scientists that all disreputable kinds of epilepsy genes and large number of variations epilepsy and hospital epilepsies are very much linked to SCN1A **[48].**

Nearly 10 % patients of GEFS+ are related with mutation in SCN1A gene and 85% patients of Dravet Syndrome are caused by mutations or removal of SCN1A. Approximately 30 SCN1A-GEFS+ mutations are recognized as yet. All missense mutations (replacement of amino acid) are there. All data is up dated regarding the functions of SCN1A but the work is continuously done to unveil the SCN1A mutations and their functions in the human body. In contrast, nearly half of above 600 SCN1A variations in Dravet Syndrome patients are due to frame shift, nonsense and splice site variations [49, 50]. These mutations are present in a specific number too early in order of SCN1A, tough illuminating that after the changed allele a ornamental protein invention is a yield, or that show of the malformed allele is reduced, declarative of hap-lo-insufficiency of SCN1A [51, 52].

Common observation it is seen commonly that epilepsy in human beings is due to mutations in three of the genes that are coding for Na-channel α subunit, which are primarily presented in CNS. Many of the idiopathic forms of all the generalized epilepsies outcomes of mutation in SCN1A gene (Nav1.1) and SCN2A gene (Nav1.2). Genomic epilepsy as well as febrile seizures plus (GEFS+; MIM 604223) mostly in the outcomes of mutation in the two above said genes [53-56] SCN1B gene with mutation that encode for Na-channel β 1 subunit [57, 58] and 2 GABAA receptors genes: GABRG2 encodes for γ 2 subunit [59-61] and GABRD encodes for γ subunit [62] also causing for epilepsy syndrome. SCN1A mutations are the fundamental the basic cause of both back breaking early years epilepsy concluded tonic-colonic seizures generalized seizures, similarly named as severely generalized epilepsy with unknown reason of infancy also recognized as Dravet syndrome (DS, MIM 607208) [40], [63].

A few patients of DS and non-threatening inherited neonatal seizures (BFNIS; MIM 607745) are caused by SCN2A mutations [64-66], and it admitted that SCN1B mutations are responsible for painful Dravet syndrome [67]. The mutation in SCN3A is causing focal epilepsy is seen in some studies (Nav1.3) [68]. The mutations in SCN9A are admitted with febrile seizures in several years ago interestingly. SCN9A gene is harmonious to be linked largely in the PNS, SCN9A may work as a genomic modification of Dravet syndrome [69]. Gene Structure

SCN1A spans approximately 84 Mb of genomic DNA and has a transcript of 8,100 bp (reference sequence NM-006920.4). The gene comprises 26 exons that encode a protein of 1,998 amino acid residues (reference sequence NP-008851.3). Splicing variability has been reported. For a detailed summary of gene and protein information, 1.



Figure 2: Spectrum of clinical phenotypes in SCN1A gene.



Figure 3: Domains and subunits of SCN1A gene responsible for epilepsy phenotypes.

Table 1:	SCN1A-related	seizure disord	ders genes and	databases.

Gene	Chromosome Locus	Protein	Locus Specific	HGMD
SCN1A	2q24.3	Sodium channel protein type 1 subunit alpha	Familial Hemiplegic Migraine (FHM) Variation Database (SCN1A)	SCN1A

SCN1A Gene Conformation

The SCN1A contains eighty-four Mb of genomic DNA approximately and its transcription comprises of eighty-one hundred base pairs (reference sequence NM_006920.4). SCN1A gen has 26 exons which are encoding for the construction of 1,998 AA remainders long protein (reference sequence NP_008851.3). Splicing mutation is described in some studies [51].

A several number of missense mutations in SCN1A gene are accepted in DS cases. Due to several missense mutations there may also halt the character of channel, may be altering the physical features of the channel, relationships with new particles or shifting or subcellular localization. Approximately all DS mutation are de novo in pediatric patients, while in the affected families with GEFS+ mutations are not as in DS patients. It is mentioned in the recent studies that almost all de novo mutations repetitively start from the fatherly chromosome **[70].**

The comparison between various cycles of mitoses in the development of spermatozoa with the development of ova and disposition of the sperm cell's DNA with methyl group to acquire mutations are able to prove the above statement. But most of the DS cases hold from beginning of SCN1A variations, approximately instances have been described them the mutation was inborn and from a slightly affected or without symptoms parental was genetic. In a large number of patients having germ line and somatic SCN1A mosaicism is described **[71-74].**

Authentication of SCN1A Mutation

Firstly, to regulate whether there are any predictable useful significances connected with alteration / mutation and secondly, to attempt feature this changed role to the detected clinical signs and symptoms of epilepsy are main goals of useful authentication of SCN1A gene modifications. This will come to be clear that although for a several number of variations of SCN1A for the 1st goalmouth that has been completed, the 2nd probably more energetic goal has verified difficult to achieve. Here is no comprehensive mechanism attainable that can explain how the level of supposed useful properties linked with epilepsy agreed with cases possessing this nutation to date. Special effects exposed by human SCN1A mutations effects on human being are shown on ion channel properties were mostly described by presenting them in extra derivative of human Na-channel subunits gene which may be described beyond of β in addition subunits. In rat, it is expressed in Xenopus oocyte beyond of β as additional subunits [75-78].

Although, the adjacent demo of appearance of changed human SCN1A derived in neural tissue [79]. In the cloning of human beings, the arrangement of SCN1A is mildly changed, 3 generalized epilepsy with febrile seizures plus variations takes place as well as extra β subunits, in other mammalian cells were completed by themselves.

The mutations are R1648H and T875M **[48]** and W1204R **[50]**. Epileptogenesis is implication for fundamental mechanism which is derived from the

presentation of these mutations is damage of functions at minor level along with continuous extra activity of neurons in the excited state and internal Na- current. The situation become more complicated while studying extra 5 mutations (four GEFS+ and one SMEI) [80]. Two GEFS+ patients (11656M of [51] and R1657C, (which had not been formerly reported), they recognized changed gating qualities of an operational channel. The qualities are understood in three previous evaluated mutations of there is no result on Na-currents or disability [79]. In difference, the A1685V GEFS+ allele of, the V 1353LGEFS+ allele of [81], and the L986F SMEI allele of [82] displayed whole deficiency of function [80]. Accordingly, as considered in this appearance scheme, only disability of SCN1A mutations is not accrediting for the clinical difference amongst SMEI and GEFS+ [83].

They assumed multiple associations of the biophysical possessions of channel with genetic modifier and metabolic, that is only channel characteristics performed not be a predictor of phenotype. SCN1A gene is very sensitive regarding functions in the atmosphere of cell disadvantaged for the studies. In the discussions of the earliest studies, their oocyte and HEK293 expression is used as a proof of SCN1A production channel with different degrees of sensitivity to β extra subunits unlike dynamic properties.

Useful level of channels is summarized by the alterations in transcription or shifting can be able to resolve these changed functional influences and display a basic key in relating mutations with the phenotype of the patient, in the membrane of cells. Further means to incorporation the extensive range of useful effects to a general phenotype are reserved together with a particular representative study by [84] who used computer for reproducing to an collection of mutation with several efficient effects. There is a validation that a variety of mutation, all of one with a miscellaneous degree of useful variations can be all yield a hypothetical increase in cellular volatility as strong-minded by an enhanced speediness of action potential gunfire by means of a standard.

Instead of any infrequent variant directly linked to the actual mutation will continually isolate with the sickness, which is not related to their importance of its functional. Exclusion of the expected mutation in large cohort of families is much important than when observed in smaller cohort families, but, in infrequent size families liable alleles with smaller significances will not ever be seen isolating. In the condition in which the mutation expected to having a pathogenic result changes an AA at a site in the protein conserved by development, or a site conserved into proteins encoding the alike gene family, this will be a strong incidental proof that the mutation is pathogenic. The most valuable condition for decision of pathogenicity was created on evolutionary conservation that distributes the strong provisional proof, earlier to the duplicating of human appearance conceptions of SCN1A [69].

DIAGNOSIS

The identification of disease is intensely depending upon bodily investigation of patient and clinical history of disease. It is very problematic to discriminate between epilepsy and seizures. The start of the first seizure having significant values in the diagnosis of disease. Info's regarding starting time of seizure, promoting factors of seizures and circumstances should be composed. Epilepsy is often not diagnosed. Thus the mandatory diagnosis is differential diagnosis e.g., arrhythmias, nonepileptic seizures, and unconsciousness. Best investigation to diagnose the epilepsy is EEG, but patients having normal EEG are not safe from epilepsy. EEG can give adequate information of classification of epilepsies. Ictal EEG results adding further information in the diagnosis of epilepsy. MRI is another helping investigation to evaluate the epilepsy. It is providing better help to the diagnosis of epilepsy.

TREATMENT

We can get rid of genetic epilepsy through rule out the genetic cause also. For evaluation of the cause we collect blood samples of epileptic patients. After extraction of DNA sequencing is done to identify the causative mutations SCN1A gene mutations are notorious for genetic epilepsy. We can see the SCN1A gene mutations in patients with family history of epilepsy. Treatments of epilepsy are antiepileptic drugs (AEDs) are commonly given. These treatments are used for long durations with maximum side effects. There is a need of proper care at efficient level.

CONCLUSION

There is enormous advancement in the field of molecular genetics. Ion channel genes belong to the gene family that is most frequently affected and mutated causing a number of brain disorders including epilepsy. Studies on the pathophysiology and mechanisms of their channelopathies might open new corridors towards better treatment options, which may be helpful in identifying the exact defects. Along with this, the advancement in the knowledge about mutated genes can help in designing of new drugs targeted to specific pathogenic mechanisms.

REFERENCES

- Forsgren L, Edvinsson SO, Nyström L, Blomquist HK. Influence of epilepsy on mortality in mental retardation: an epidemiologic study. Epilepsia. 1996 Oct;37(10):956-63.
- 2. Hesdorffer DC, Caplan R, Berg AT. Familial clustering of epilepsy and behavioral disorders: evidence for a shared genetic basis. Epilepsia. 2012 Feb;53(2):301-7.
- **3.** Pandolfo M. Genetics of epilepsy. Semin Neurol. 2011;31,506-18.
- **4.** Steinlein OK. Channelopathies can cause epilepsy in man. European Journal of Pain. 2002 Jan 1; 6:27-34.
- Hauser WA, Annegers JF, Kurland LT. Incidence of epilepsy and unprovoked seizures in Rochester, Minnesota: 1935–1984. Epilepsia. 1993 May;34(3):453-8.
- 6. Brodie MJ, Elder AT & Kwan P. Epilepsy in later life. The lancet Neurology. (2019); 8, 1019-1030.
- 7. Newton CR, Garcia HH. Epilepsy in poor regions of the world. The Lancet. 2012 Sep 29;380(9848):1193-201.
- Eraky MA, Abdel-Hady S, Abdallah KF. Seropositivity of Toxoplasma gondii and Toxocara spp. in children with cryptogenic epilepsy, Benha, Egypt. The Korean Journal of Parasitology. 2016 Jun;54(3):335.
- Abubakar II, Tillmann T, Banerjee A. Global, regional, and national age-sex specific all-cause and cause-specific mortality for 240 causes of death, 1990-2013: a systematic analysis for the Global Burden of Disease Study 2013. Lancet. 2015 Jan 10;385(9963):117-71.
- **10.** Liu S, Zheng P. Altered PKA modulation in the Nav1. 1 epilepsy variant 11656M. Journal of Neurophysiology. 2013 Nov 1;110(9):2090-8.
- **11.** Scheffer IE, Zhang YH, Jansen FE, Dibbens L. Dravet syndrome or genetic (generalized) epilepsy with febrile seizures plus? Brain and Development. 2009 May 1;31(5):394-400.
- Scheffer IE, Berkovic SF. Generalized epilepsy with febrile seizures plus. A genetic disorder with heterogeneous clinical phenotypes. Brain: a journal of neurology. 1997 Mar 1;120(3):479-90.
- **13.** Mullen SA, Scheffer IE. Translational research in epilepsy genetics: sodium channels in man to interneuronopathy in mouse. Archives of neurology. 2009 Jan 1;66(1):21-6.
- **14.** Srinivasan J, Wallace KA, Scheffer IE. Febrile seizures. Australian family physician. 2005 Dec;34(12).
- Selmer KK, Eriksson AS, Brandal K, Egeland T, Tallaksen C, Undlien DE. Parental SCN1A mutation mosaicism in familial Dravet syndrome. Clinical genetics. 2009 Oct;76(4):398-403.
- Granata T. Comprehensive care of children with Dravet syndrome. Epilepsia. 2011 Apr; 52:90-4.
- England MJ, Liverman CT, Schultz AM, Strawbridge LM. Epilepsy across the spectrum: Promoting health and understanding.: A summary of the Institute of Medicine report. Epilepsy & Behavior. 2012 Oct 1;25(2):266-76.
- **18.** Eadie MJ. Shortcomings in the current treatment of epilepsy. Expert review of neurotherapeutics. 2012 Dec 1;12(12):1419-27.
- **19.** Noebels JL. The biology of epilepsy genes. Annual review of neuroscience. 2003 Mar;26(1):599-625.
- **20.** Catterall WA. From ionic currents to molecular mechanisms: the structure and function of voltage-gated sodium channels. Neuron. 2000 Apr 1;26(1):13-25.
- **21.** Isom LL, De Jongh KS, Catterall WA. Auxiliary subunits of voltage-gated ion channels. Neuron. 1994 Jun 1;12(6):1183-94.
- 22. Morgan K, Stevens EB, Shah B, Cox PJ, Dixon AK, Lee K, Pinnock RD, Hughes J, Richardson PJ, Mizuguchi K, Jackson AP. β3: an additional auxiliary subunit of the voltage-sensitive sodium channel that modulates channel gating with distinct kinetics. Proceedings of the National Academy of Sciences. 2000 Feb 29;97(5):2308-13.

- 23. Yu FH, Westenbroek RE, Silos-Santiago I, McCormick KA, Lawson D, Ge P, Ferriera H, Lilly J, DiStefano PS, Catterall WA, Scheuer T. Sodium channel beta4, a new disulfide-linked auxiliary subunit with similarity to beta2. The Journal of neuroscience: the official journal of the Society for Neuroscience. 2003 Aug 1;23(20):7577-85.
- **24.** Trimmer JS, Rhodes KJ. Localization of voltage-gated ion channels in mammalian brain. Annual review of physiology. 2004; 66:477.
- **25.** Beckh S, Noda M, Lübbert H, Numa S. Differential regulation of three sodium channel messenger RNAs in the rat central nervous system during development. The EMBO journal. 1989 Dec;8(12):3611-6.
- 26. Waxman SG. The neuron as a dynamic electrogenic machine: modulation of sodium–channel expression as a basis for functional plasticity in neurons. Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences. 2000 Feb 29;355(1394):199-213.
- 27. Keller DI, Huang H, Zhao J, Frank R, Suarez V, Delacrétaz E, Brink M, Osswald S, Schwick N, Chahine M. A novel SCN5A mutation, F1344S, identified in a patient with Brugada syndrome and fever-induced ventricular fibrillation. Cardiovascular research. 2006 Jun 1;70(3):521-9.
- 28. Fertleman CR, Baker MD, Parker KA, Moffatt S, Elmslie FV, Abrahamsen B, Ostman J, Klugbauer N, Wood JN, Gardiner RM, Rees M. SCN9A mutations in paroxysmal extreme pain disorder: allelic variants underlie distinct channel defects and phenotypes. Neuron. 2006 Dec 7;52(5):767-74.
- Whitaker WR, Faull RL, Waldvogel HJ, Plumpton CJ, Emson PC, Clare JJ. Comparative distribution of voltagegated sodium channel proteins in human brain. Molecular Brain Research. 2001 Mar 31;88(1-2):37-53.
- 30. Black JA, Cummins TR, Plumpton C, Chen YH, Hormuzdiar W, Clare JJ, Waxman SG. Upregulation of a silent sodium channel after peripheral, but not central, nerve injury in DRG neurons. Journal of neurophysiology. 1999 Nov 1;82(5):2776-85.
- **31.** Hains BC, Klein JP, Saab CY, Craner MJ, Black JA, Waxman SG. Upregulation of sodium channel Nav1. 3 and functional involvement in neuronal hyperexcitability associated with central neuropathic pain after spinal cord injury. Journal of Neuroscience. 2003 Oct 1;23(26):8881-92.
- **32.** Yu FH, Mantegazza M, Westenbroek RE, Robbins CA, Kalume F, Burton KA, Spain WJ, McKnight GS, Scheuer T, Catterall WA. Reduced sodium current in GABAergic interneurons in a mouse model of severe myoclonic epilepsy in infancy. Nature neuroscience. 2006 Sep;9(9):1142-9.
- 33. Guo F, Yu N, Cai JQ, Quinn T, Zong ZH, Zeng YJ, Hao LY. Voltage-gated sodium channel Nav1. 1, Nav1. 3 and β1 subunit were up-regulated in the hippocampus of spontaneously epileptic rat. Brain research bulletin. 2008 Jan 31;75(1):179-87.
- 34. Felts PA, Yokoyama S, Dib-Hajj S, Black JA, Waxman SG. Sodium channel α-subunit mRNAs I, II, III, NaG, Na6 and hNE (PN1): different expression patterns in developing rat nervous system. Molecular brain research. 1997 Apr 1;45(1):71-82.
- **35.** Schaller KL, Krzemien DM, Yarowsky PJ, Krueger BK, Caldwell JH. A novel, abundant sodium channel expressed in neurons and glia. Journal of Neuroscience. 1995 May 1;15(5):3231-42.
- 36. Gordon D, Merrick D, Auld V, Dunn R, Goldin AL, Davidson N, Catterall WA. Tissue-specific expression of the RI and RII sodium channel subtypes. Proceedings of the National Academy of Sciences. 1987 Dec;84(23):8682-6.

- Westenbroek RE, Merrick DK, Catterall WA. Differential subcellular localization of the RI and RII Na+ channel subtypes in central neurons. Neuron. 1989 Dec 1;3(6):695-704.
- 38. Gong B, Rhodes KJ, Bekele-Arcuri Z, Trimmer JS. Type I and type II Na⁺ channel α-subunit polypeptides exhibit distinct spatial and temporal patterning, and association with auxiliary subunits in rat brain. Journal of Comparative Neurology. 1999 Sep 20;412(2):342-52.
- **39.** Ogiwara I, Miyamoto H, Morita N, Atapour N, Mazaki E, Inoue I, Takeuchi T, Itohara S, Yanagawa Y, Obata K, Furuichi T. Nav1. 1 localizes to axons of parvalbuminpositive inhibitory interneurons: a circuit basis for epileptic seizures in mice carrying an Scn1a gene mutation. Journal of Neuroscience. 2007 May 30;27(22):5903-14.
- 40. Whitaker W, Faull R, Waldvogel H, Plumpton C, Burbidge S, Emson P, Clare J. Localization of the type VI voltage-gated sodium channel protein in human CNS. Neuroreport. 1999 Nov 26;10(17):3703-9.
- Tzoumaka E, Tischler AC, Sangameswaran L, Eglen RM, Hunter JC, Novakovic SD. Differential distribution of the tetrodotoxin-sensitive rPN4/NaCh6/Scn8a sodium channel in the nervous system. Journal of neuroscience research. 2000 Apr 1;60(1):37-44.
- 42. Caldwell JH, Schaller KL, Lasher RS, Peles E, Levinson SR. Sodium channel Nav1. 6 is localized at nodes of Ranvier, dendrites, and synapses. Proceedings of the National Academy of Sciences. 2000 May 9;97(10):5616-20.
- Krzemien DM, Schaller KL, Levinson SR, Caldwell JH. Immunolocalization of sodium channel isoform NaCh6 in the nervous system. Journal of Comparative Neurology. 2000 Apr 24;420(1):70-83.
- **44.** Hu W, Tian C, Li T, Yang M, Hou H, Shu Y. Distinct contributions of Nav1. 6 and Nav1. 2 in action potential initiation and backpropagation. Nature neuroscience. 2009 Aug;12(8):996-1002.
- Mulley JC, Scheffer IE, Petrou S, Dibbens LM, Berkovic SF, Harkin LA. SCN1A mutations and epilepsy. Human mutation. 2005 Jun;25(6):535-42.
- 46. Ceulemans BP, Claes LR, Lagae LG. Clinical correlations of mutations in the SCN1A gene: from febrile seizures to severe myoclonic epilepsy in infancy. Pediatric neurology. 2004 Apr 1;30(4):236-43.
- 47. Mulley JC, Nelson P, Guerrero S, Dibbens L, Iona X, McMahon JM, Harkin L, Schouten J, Yu S, Berkovic SF, Scheffer IE. A new molecular mechanism for severe myoclonic epilepsy of infancy: exonic deletions in SCN1A. Neurology. 2006 Sep 26;67(6):1094-5.
- 48. Berkovic SF, Harkin L, McMahon JM, Pelekanos JT, Zuberi SM, Wirrell EC, Gill DS, Iona X, Mulley JC, Scheffer IE. De-novo mutations of the sodium channel gene SCN1A in alleged vaccine encephalopathy: a retrospective study. The Lancet Neurology. 2006 Jun 1;5(6):488-92.
- **49.** Lossin C. A catalog of SCN1A variants. Brain and Development. 2009 Feb 1;31(2):114-30.
- 50. Claes LR, Deprez L, Suls A, Baets J, Smets K, Van Dyck T, Deconinck T, Jordanova A, De Jonghe P. The SCN1A variant database: a novel research and diagnostic tool. Human mutation. 2009 Oct;30(10): E904-20.
- Meisler MH, Kearney JA. Sodium channel mutations in epilepsy and other neurological disorders. The Journal of clinical investigation. 2005 Aug 1;115(8):2010-7.
- Catterall WA, Dib-Hajj S, Meisler MH, Pietrobon D. Inherited neuronal ion channelopathies: new windows on complex neurological diseases. Journal of Neuroscience. 2008 Nov 12;28(46):11768-77.
- Escayg A, MacDonald BT, Meisler MH, Baulac S, Huberfeld G, An-Gourfinkel I, Brice A, LeGuern E, Moulard B, Chaigne D, Buresi C. Mutations of SCN1A,

encoding a neuronal sodium channel, in two families with GEFS+ 2. Nature genetics. 2000 Apr;24(4):343-5.

- 54. Sugawara T, Tsurubuchi Y, Agarwala KL, Ito M, Fukuma G, Mazaki-Miyazaki E, Nagafuji H, Noda M, Imoto K, Wada K, Mitsudome A. A missense mutation of the Na+ channel αII subunit gene Na v 1.2 in a patient with febrile and afebrile seizures causes channel dysfunction. Proceedings of the National Academy of Sciences. 2001 May 22;98(11):6384-9.
- 55. Escayg A, Heils A, MacDonald BT, Haug K, Sander T, Meisler MH. A novel SCN1A mutation associated with generalized epilepsy with febrile seizures plus—and prevalence of variants in patients with epilepsy. The American Journal of Human Genetics. 2001 Apr 1;68(4):866-73.
- Mulley JC, Scheffer IE, Petrou S, Dibbens LM, Berkovic SF, Harkin LA. SCN1A mutations and epilepsy. Human mutation. 2005 Jun;25(6):535-42.
- 57. Wallace RH, Wang DW, Singh R, Scheffer IE, George AL, Phillips HA, Saar K, Reis A, Johnson EW, Sutherland GR, Berkovic SF. Febrile seizures and generalized epilepsy associated with a mutation in the Na+-channel ß1 subunit gene SCN1B. Nature genetics. 1998 Aug;19(4):366-70.
- 58. Wallace RH, Scheffer IE, Parasivam G, Barnett S, Wallace GB, Sutherland GR, Berkovic SF, Mulley JC. Generalized epilepsy with febrile seizures plus: mutation of the sodium channel subunit SCN1B. Neurology. 2002 May 14;58(9):1426-9.
- 59. Baulac S, Huberfeld G, Gourfinkel-An I, Mitropoulou G, Beranger A, Prud'homme JF, Baulac M, Brice A, Bruzzone R, LeGuern E. First genetic evidence of GABAA receptor dysfunction in epilepsy: a mutation in the γ2-subunit gene. Nature genetics. 2001 May;28(1):46-8.
- 60. Wallace RH, Marini C, Petrou S, Harkin LA, Bowser DN, Panchal RG, Williams DA, Sutherland GR, Mulley JC, Scheffer IE, Berkovic SF. Mutant GABAA receptor γ2subunit in childhood absence epilepsy and febrile seizures. Nature genetics. 2001 May;28(1):49-52.
- 61. Harkin LA, Bowser DN, Dibbens LM, Singh R, Phillips F, Wallace RH, Richards MC, Williams DA, Mulley JC, Berkovic SF, Scheffer IE. Truncation of the GABAAreceptor γ2 subunit in a family with generalized epilepsy with febrile seizures plus. The American Journal of Human Genetics. 2002 Feb 1;70(2):530-6.
- 62. Dibbens LM, Feng HJ, Richards MC, Harkin LA, Hodgson BL, Scott D, Jenkins M, Petrou S, Sutherland GR, Scheffer IE, Berkovic SF. GABRD encoding a protein for extra-or peri-synaptic GABAA receptors is a susceptibility locus for generalized epilepsies. Human molecular genetics. 2004 Jul 1;13(13):1315-9.
- **63.** Fujiwara T. Clinical spectrum of mutations in SCN1A gene: severe myoclonic epilepsy in infancy and related epilepsies. Epilepsy research. 2006 Aug 1; 70:223-30.
- 64. Kamiya K, Kaneda M, Sugawara T, Mazaki E, Okamura N, Montal M, Makita N, Tanaka M, Fukushima K, Fujiwara T, Inoue Y. A nonsense mutation of the sodium channel gene SCN2A in a patient with intractable epilepsy and mental decline. Journal of Neuroscience. 2004 Mar 17;24(11):2690-8.
- 65. Ogiwara I, Ito K, Sawaishi Y, Osaka H, Mazaki E, Inoue I, Montal M, Hashikawa T, Shike T, Fujiwara T, Inoue Y. De novo mutations of voltage-gated sodium channel αII gene SCN2A in intractable epilepsies. Neurology. 2009 Sep 29;73(13):1046-53.
- 66. Shi X, Yasumoto S, Nakagawa E, Fukasawa T, Uchiya S, Hirose S. Missense mutation of the sodium channel gene SCN2A causes Dravet syndrome. Brain and Development. 2009 Nov 1;31(10):758-62.
- **67.** Patino GA, Claes LR, Lopez-Santiago LF. Sfilata E, Dondeti RSR, Chen C, O'Malley a H, Gray CBB, Miyazaki

H, Nukina N, Oyama F, De Jonghe P, Isom LL. A functional null mutation of SCN1B in a patient with Dravet syndrome. J Neurosci. 2009; 29:10764-78.

- **68.** Holland KD, Kearney JA, Glauser TA, Buck G, Keddache M, Blankston JR, Glaaser IW, Kass RS, Meisler MH. Mutation of sodium channel SCN3A in a patient with cryptogenic pediatric partial epilepsy. Neuroscience letters. 2008 Mar 5;433(1):65-70.
- **69.** Singh NA, Pappas C, Dahle EJ, Claes LR, Pruess TH, De Jonghe P, Thompson J, Dixon M, Gurnett C, Peiffer A, White HS. A role of SCN9A in human epilepsies, as a cause of febrile seizures and as a potential modifier of Dravet syndrome. PLoS genetics. 2009 Sep 18;5(9): e1000649.
- **70.** Gardiner M. Genetics of idiopathic generalized epilepsies. Epilepsia. 2005 Nov; 46:15-20.
- **71.** Heron SE, Scheffer IE, Iona X, Zuberi SM, Birch R, McMahon JM, Bruce CM, Berkovic SF, Mulley JC. De novo SCN1A mutations in Dravet syndrome and related epileptic encephalopathies are largely of paternal origin. Journal of medical genetics. 2010 Feb 1;47(2):137-41.
- **72.** Gennaro E, Santorelli FM, Bertini E, Buti D, Gaggero R, Gobbi G, Lini M, Granata T, Freri E, Parmeggiani A, Striano P. Somatic and germline mosaicisms in severe myoclonic epilepsy of infancy. Biochemical and biophysical research communications. 2006 Mar 10;341(2):489-93.
- **73.** Steinlein OK. Mechanisms underlying epilepsies associated with sodium channel mutations. Progress in Brain Research. 2014 Jan 1; 213:97-111.
- 74. Depienne C, Arzimanoglou A, Trouillard O, Fedirko E, Baulac S, Saint-Martin C, Ruberg M, Dravet C, Nabbout R, Baulac M, Gourfinkel-An I. Parental mosaicism can cause recurrent transmission of SCN1A mutations associated with severe myoclonic epilepsy of infancy. Human mutation. 2006 Apr;27(4):389-.
- Marini C, Mei D, Helen Cross J, Guerrini R. Mosaic SCN1A mutation in familial severe myoclonic epilepsy of infancy. Epilepsia. 2006 Oct;47(10):1737-40.
- Alekov AK, Rahman MM, Mitrovic N, Lehmann-Horn F, Lerche H. Enhanced inactivation and acceleration of

activation of the sodium channel associated with epilepsy in man. European Journal of Neuroscience. 2001 Jun;13(11):2171-6.

- Alekov AK, Rahman MM, Mitrovic N, LehmannHorn F, Lerche H. A sodium channel mutation causing epilepsy in man exhibits subtle defects in fast inactivation and activation in vitro. The Journal of Physiology. 2000 Dec;529(3):533-40.
- 78. Spampanato J, Escayg A, Meisler MH, Goldin AL. Generalized epilepsy with febrile seizures plus type 2 mutation W1204R alters voltage-dependent gating of Nav1. 1 sodium channels. Neuroscience. 2003 Jan 15;116(1):37-48.
- 79. Spampanato J, Escayg A, Meisler MH, Goldin AL. Functional effects of two voltage-gated sodium channel mutations that cause generalized epilepsy with febrile seizures plus type 2. Journal of neuroscience. 2001 Oct 1:21(19):7481-90.
- Lossin, C., Wang, D. W., Rhodes, T. H., Vanoye, C. G. & George, A. L., Jr. (2002). Molecular basis of an inherited epilepsy. Neuron 34, 877-84.
- Lossin C, Rhodes TH, Desai RR, Vanoye CG, Wang D, Carniciu S, Devinsky O, George AL. Epilepsy-associated dysfunction in the voltage-gated neuronal sodium channel SCN1A. Journal of Neuroscience. 2003 Dec 10;23(36):11289-95.
- 82. Sugawara T, Mazaki–Miyazaki E, Ito M, Nagafuji H, Fukuma G, Mitsudome A, Wada K, Kaneko S, Hirose S, Yamakawa K. Nav1. 1 mutations cause febrile seizures associated with afebrile partial seizures. Neurology. 2001 Aug 28;57(4):703-5.
- 83. Claes L, Del-Favero J, Ceulemans B, Lagae L, Van Broeckhoven C, De Jonghe P. De novo mutations in the sodium-channel gene SCN1A cause severe myoclonic epilepsy of infancy. The American Journal of Human Genetics. 2001 Jun 1;68(6):1327-32.
- 84. Sugawara T, Tsurubuchi Y, Fujiwara T, Mazaki-Miyazaki E, Nagata K, Montal M, Inoue Y, Yamakawa K. Nav1. 1 channels with mutations of severe myoclonic epilepsy in infancy display attenuated currents. Epilepsy research. 2003 May 1;54(2-3):201-7.